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### **The Role of Phosphoinositides in the Interaction of Myelin Basic Protein with the Oligodendroglial Cell Membrane**

The only protein known to be essential for myelin formation and compaction in the central nervous system is myelin basic protein (MBP). Oligodendrocytes in mutant mice that lack MBP-expression are unable to deposit a functional myelin sheath and the few lamellae formed around axons are not compacted. The association of MBP as a positively-charged protein with negatively charged membranes is therefore crucial for myelination, but the mechanisms by which MBP associates with the myelin membrane remains elusive. In this study, I demonstrate that the signaling lipid phosphatidylinositol (4,5)-bisphosphate (PIP<sub>2</sub>) is important for the stable association of MBP with cellular membranes. This association is lost upon specific reduction of PIP<sub>2</sub> levels following the overexpression of a phosphoinositide-5-phosphatase (Synaptojanin 1) that selectively hydrolyzes PIP<sub>2</sub>. The association is also lost through elevated intracellular Ca<sup>2+</sup> levels. Moreover, since MBP interacts with the membranes electrostatically, the experimental decrease of membrane charges at the intracellular membrane surface (through PIP<sub>2</sub> dephosphorylation) was shown to cause the dissociation of MBP from the plasma membrane. Experiments presented here further implicate that one putative PIP<sub>2</sub> binding domain of MBP lies within the exon-1 encoded region. The relevance of this protein-lipid interaction was demonstrated for the corpus callosum of mice, analyzed by electron microscopy after reducing membrane surface charges in acute brain slices. Here, PIP<sub>2</sub> hydrolysis led to the loss of myelin compaction. A related phosphoinositide that might play a role in myelin formation is the signaling lipid phosphatidylinositol(3,4,5)-trisphosphate (PIP<sub>3</sub>). We found PIP<sub>3</sub> and some downstream polarizing factors to be accumulated at the tips of growing cellular processes in both immortalized and primary oligodendrocytes. Taken together, the results presented here demonstrate that PIP<sub>2</sub> and PIP<sub>3</sub> play an important role in MBP association to the plasma membrane and oligodendroglial polarity. This association might induce the formation of lipid clusters, which could serve as a signalling platform for polarization of oligodendrocytes through PIP<sub>3</sub> signalling. These findings provide a novel link between phosphoinositol metabolism and MBP function in oligodendrocytes in development and disease.